

New species of the *Pseudancistrus barbatus* group (Siluriformes, Loricariidae) with comments on its biogeography and dispersal routes

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Abstract

A new species of *Pseudancistrus* is described from the Tapajós Basin, and assigned to the *P. barbatus* group by having hypertrophied odontodes along the snout and lacking evertible cheek plates. The new species is distinguished from other species in that group (*P. barbatus*, *P. corantijnensis*, *P. depressus* and *P. nigrescens*) by its pattern of spots, length and color of snout odontodes, greater head depth, cleithral width, anal-fin spine length, peduncle depth and internares width. Molecular phylogenetic results corroborate placement of the new species in the *Pseudancistrus barbatus* group which is otherwise distributed in the Xingu Basin and rivers draining the Guyana Shield into the Atlantic Ocean. Topology tests strongly reject alternative hypotheses supporting close relationships with *Guyanancistrus*, *Lithoxancistrus* or the species *Pseudancistrus pectegenitor*, *P. sidereus* and *P. genisetiger*. Additionally, we propose two hypotheses on the distribution of the new species in the rio Tapajós, a Brazilian Shield drainage. The first one proposes that ancestral stock of the *P. barbatus* group was widely distributed throughout rivers draining the Guyana and Brazilian

shields, and the species *P. zawadzki* and *Pseudancistrus* sp. L17 are in the limit of the distribution for the group in Tapajós and Xingu rivers. The second hypothesis proposes that ancestral stock of the *P. barbatus* group was restricted to Guyana Shield rivers, and that headwater capture events permitted several dispersal routes through Guyana and Amazon rivers, permitted that the ancestral lineages of *Pseudancistrus* sp. L17 and *P. zawadzki* reached the rivers of Amazon basin.

Keywords

Ancistrini, freshwater, molecular phylogeny, F-reticulon 4, Brazilian Shield

Introduction

Ancistrini is a highly diverse tribe of the subfamily Hypostominae, with 30 genera (Lujan and Armbruster 2011; Covain and Fisch-Muller 2012; Salcedo 2013) and 252 valid species (Eschmeyer and Fong 2013) widely distributed in the Neotropics from rivers in Panamá to the La Plata system in Argentina. Armbruster (2004a) provided morphological support for the monophyly of Ancistrini based on his extensive analysis of relationships within Loricariidae. Molecular data, however, suggested that Ancistrini is not monophyletic (Montoya-Burgos 1998; Covain and Fisch-Muller 2012).

Species of the genus *Pseudancistrus* Bleeker, 1862 are distributed in the Orinoco, Amazon and Jaguaribe river systems, and rivers draining the Guyana Shield into the Atlantic Ocean. Armbruster (2004a) recognized *Pseudancistrus* as a monophyletic group and included *Guyanancistrus* Isbrücker, Seidel, Michels, Schraml & Werner, 2001 and *Lithoxancistrus* Isbrücker, Nijssen & Cala, 1988 in its synonymy. Based on molecular and morphological data, Chambrier and Montoya-Burgos (2008) defined a subgroup within *Pseudancistrus* called the *Pseudancistrus barbatus* group and composed of *P. barbatus* (Valenciennes, 1840), *P. depressus* (Günther, 1868), *P. nigrescens* Eigenmann, 1912, and *P. corantijniensis* De Chambrier & Montoya-Burgos, 2008. That group was morphologically defined by having hypertrophied odontodes along the snout and lacking evertible cheek plates. Recently, Covain and Fisch-Muller (2012) suggested that *Pseudancistrus guentheri* (Regan, 1904) and *P. kwinti* Willink, Mol & Chernoff, 2010 may be added to the *P. barbatus* group. Covain and Fisch-Muller (2012) also recognized *Pseudancistrus* as paraphyletic, and restricted the genus by the *P. barbatus* group. They revalidated the genera *Guyanancistrus* and *Lithoxancistrus*, and considered *P. pectegenitor* Lujan, Armbruster & Sabaj Pérez, 2007, *P. sidereus* Armbruster, 2004b, and *P. genisetiger* Fowler, 1941 to represent two separate lineages unrelated to *Pseudancistrus*. Covain and Fisch-Muller (2012) suggested that these two lineages represent undescribed genera.

In this paper, we present a formal description of a new species of *Pseudancistrus* from the Tapajós river basin. Additionally, we provide a phylogenetic context for the new species based on analysis of sequence data of F-reticulon 4 nuclear gene, and a brief discussion of biogeographic scenarios that may explain the distribution of the new species in the rio Tapajós and northern Brazilian Shield.

Material and methods

Sampling and morphological analysis

After capture, fish were anesthetized using 1% benzocaine in water, and either preserved in 95% ethanol for molecular studies or fixed in 10% formaldehyde for morphological studies. Vouchers and tissues were deposited in the collection of the Laboratório de Biologia e Genética de Peixes (LBP) and Museu de Zoologia da Universidade de São Paulo (MZUSP), Brazil, Muséum d'histoire naturelle de la ville de Genève (MHNG), Switzerland, Academy of Natural Sciences of Philadelphia (ANSP) and Auburn University (AUM), U.S.A., and Smithsonian Tropical Research Institute (STRI), Panama. Measurements and counts were taken on left side of specimens. Measurements follow Armbruster (2003), and were taken point to point to the nearest 0.1 mm with digital calipers.

DNA sequencing

Total DNA was extracted from ethanol-preserved muscle, fin, and liver samples using the Wizard Genomic DNA Purification Kit (Promega, Madison, Wisconsin, U.S.A.). Partial sequences of F-reticulon 4 were amplified using polymerase chain reaction (PCR) with the following primers from Chiachio et al. (2008): Freticul4-D 5'-AGG CTA ACT CGC TYT SGG CTT TG-3', Freticul4-R 5'-GGC AVA GRG CRA ART CCA TCT C-3', Freticul4 D2 5'-CTT TGG TTC GGA ATG GAA AC-3', Freticul4 R2 5'-AAR TCC ATC TCA CGC AGG A-3', Freticul4 iR 5'-AGG CTC TGC AGT TTC TCT AG-3'.

Amplifications were performed in a total volume of 12.5 µl containing 1.25 µl of 10X PCR buffer (20 mM Tris-HCl, pH 8.0, 40 mM NaCl, 2 mM Sodium Phosphate, 0.1 mM EDTA, 1 mM DTT, stabilizers, 50% (v/v) glycerol), 0.375 µl MgCl₂ (50nM), 0.25 µl dNTPs (2 nM), 0.25 µl (each 5 mM primer), 0.05 µl Platinum® Taq DNA Polymerase (Invitrogen), 1 µl template DNA (50 ng), and 9.075 µl ddH₂O. The nuclear markers were amplified in two PCR experiments; the first amplification using the primers Freticul4-D and Freticul4-R for 37–40 cycles (30 sec at 95°C, 30 sec at 48°C, and 135 sec at 72°C); and the second amplification using the primers Freticul4 D2, Freticul4 R2, and Freticul4 iR for 37–40 cycles (30 sec at 95°C, 30 sec at 53–54°C, and 135 sec at 72°C).

The products were then identified on a 1% agarose gel. The PCR products were purified using ExoSap-IT® (USB, Affymetrix Corporation, Cleveland, Ohio) following the manufacturer's instructions. The purified PCR products were used to make a sequencing PCR using the BigDye™ Terminator v 3.1 Cycle Sequencing Ready Reaction Kit (Applied Biosystems- Life Technologies do Brasil Ltda, Vila Guarani, SP, Brazil). Subsequently, the amplified DNA was purified again and loaded onto a 3130-Genetic Analyzer automatic sequencer (Applied Biosystems), in the Instituto de Biociências, Universidade Estadual Paulista, Botucatu, São Paulo. Contigs were as-

sembled and edited in BioEdit 7.0.9.0 (Hall 1999). Where uncertainty of nucleotide identity was detected, IUPAC ambiguity codes were applied. All sequences obtained in this study were deposited in GenBank (Table 3).

Sequence alignment and phylogenetic analyses

The DNA sequences were aligned using ClustalW program implemented in DAMBE 5.2.31 (Xia and Xie 2001) and edited in BioEdit 7.0.1 (Hall 1999), using default parameters. The alignments were inspected by eye for any obvious misalignments that were then corrected. Alignment errors only were changed where indels of 1 bp were added to introns of the reticulon gene. The sequence of F-reticulon 4 of the new species was sequenced twice, and a preliminary phylogenetic analysis was performed to control potential sequencing errors involving pseudogenes, paralogous copies or laboratory cross-contamination or mistakes during manipulations of samples. Nucleotide variation was examined using MEGA 5.0 (Tamura et al. 2007). To evaluate the occurrence of substitution saturation, we estimated the index of substitution saturation (Iss) in DAMBE 5.2.31 (Xia and Xie 2001), as described by Xia et al. (2003) and Xia and Lemey (2009).

Maximum-Likelihood (ML) analyses were performed using RAxML Web-Servers (Randomized Accelerated Maximum Likelihood, Stamatakis et al. 2008) which implements a faster algorithm of heuristic search with bootstrap pseudoreplicates (RBS). Bootstrap resampling (Felsenstein 1985) was applied to assess support for individual nodes using 1,000 replicates. Random starting trees were used for each independent ML tree search and all other parameters were set on default values. The ML analysis was conducted under a Generalized Time Reversible (GTR) model, with Gamma distribution (G) and Invariable Sites according to Modeltest 3.7 results (Posada and Crandall 1998). Gaps were treated as missing data.

Alternative tree topologies were evaluated in the program Treefinder (Jobb et al. 2004) using the Shimodaira and Hasegawa (SH) test (Shimodaira and Hasegawa 1999), the Approximately Unbiased (AU) test (Shimodaira 2002), and the Expected Likelihood Weights (ELW) method (Strimmer and Rambaut 2002). All tests were conducted under ML with a GTR model and Gamma distribution.

Results

Pseudancistrus zawadzki sp. n.

<http://zoobank.org/F244A7A4-253A-49B8-B027-16B640FDBCCF>

http://species-id.net/wiki/Pseudancistrus_zawadzki

Figure 1, Table 1

Holotype. MZUSP 115056, male, 116.4 mm SL. Brazil: Pará State: municipality of Itaituba: rio Tapajós (Amazon basin), 04°33'09.7"S, 56°17'59.6"W, 11 June 2012, R. Britzke and CEPTA's team.



Figure 1. *Pseudancistrus zawadzki*, MZUSP 115056, holotype, male, 116.4 mm SL; Pará State, Tapajós river basin, Brazil.

Paratypes. Brazil: Pará State: municipality of Itaituba: LBP 15045 (2 females, 97.9–128.7 mm SL), LBP 17724 (1 female, 87.5 mm SL), collected with holotype; LBP 16195 (1 male, 116.4 mm SL), rio Tracua (trib. rio Tapajós), 04°28'11.2"S, 56°17'01.1"W.

Diagnosis. *Pseudancistrus zawadzki* is distinguished from all congeners, except species of the *P. barbatus* group, by presence of hypertrophied odontodes along the snout

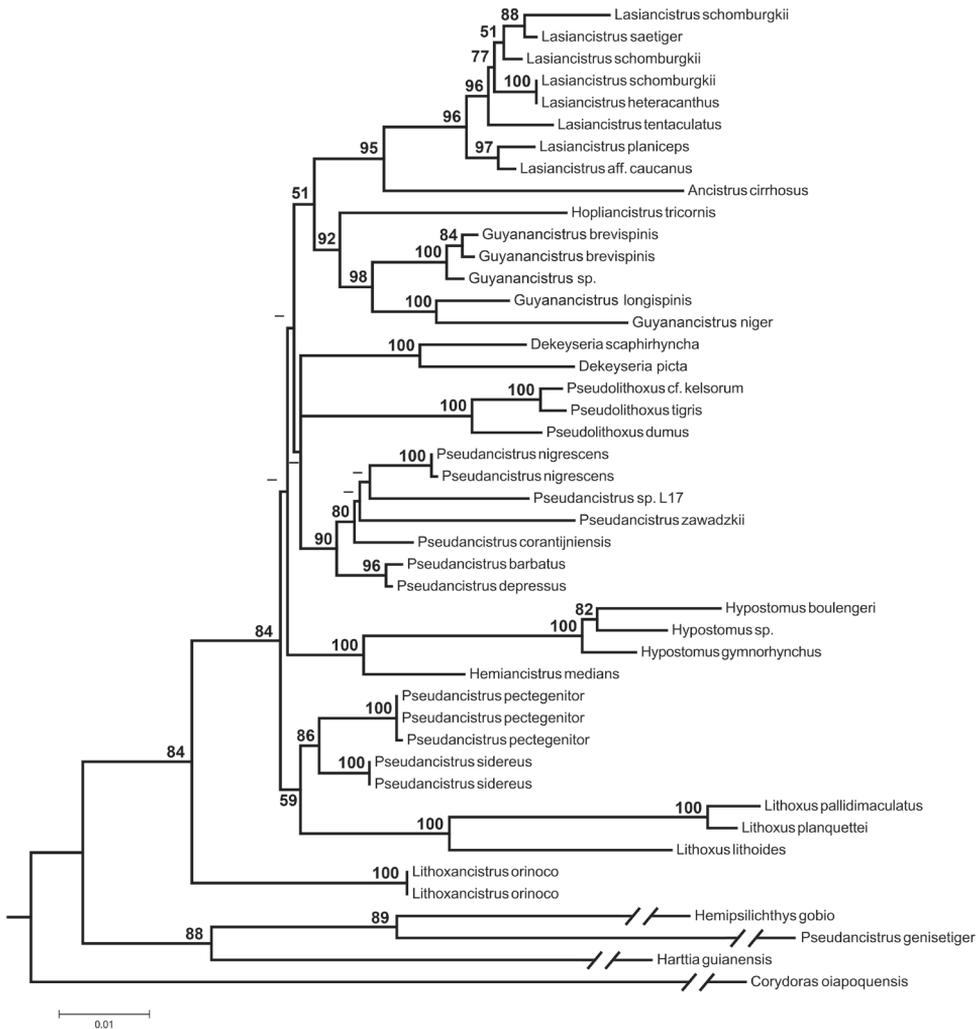


Figure 2. Maximum-likelihood tree based on nuclear gene sequence F-reticulon 4 ($-\ln L = 11470.59$). Numbers next to nodes are bootstrap values based on 1,000 pseudoreplicates. Values below 50% are not shown.

margin and the lack of evertible cheek plates. It further differs from two members of that group, *P. barbatus* and *P. depressus*, by having whitish spots that abruptly increase in size between the head (diameter 1.1–1.3 mm) and body (diameter 2.6–3.0 mm) (vs. whitish spots very small on whole body less than 1 mm), and snout odontodes yellowish (vs. snout odontodes reddish-brown). The new species differs from the other two members of the *P. barbatus* group, *P. corantijniensis* and *P. nigrescens*, by having odontodes along margin of snout increasing gradually in length from posterior of snout tip to cheek (vs. length of snout odontodes more uniform, smaller on tip of snout) and by having odontodes relatively longer on the most posterior portion of the nonevertible cheek plates (Fig. 1) (vs. odontodes shorter) (see fig. 3 in Chambrier and Montoya-

Table 1. Morphometric data for *Pseudancistrus zawadzki*.

	<i>Pseudancistrus zawadzki</i> n = 5			
	Holotype	Range	Mean	SD
Standard length (SL)	116.4	128.7–87.5	109.5	
Percents of SL				
Predorsal length	43.3	43.1–46.1	44.5	1.3
Head length	36.6	32.9–37.8	36.3	1.9
Head-dorsal length	6.7	6.7–9.2	8.1	1.2
Cleithral width	35.2	35.2–38.0	36.7	1.2
Head pectoral length	30.5	29.6–32.2	30.9	0.9
Thorax length	23.5	21.2–23.5	22.5	1.1
Pectoral-spine length	31.5	31.3–33.2	31.9	0.7
Abdominal length	24.2	22.6–26.1	24.3	1.3
Pelvic-spine length	28.4	25.6–28.4	27.2	1.2
Post-anal length	31.2	29.6–31.2	30.5	0.7
Anal-fin spine length	12.5	11.9–13.8	12.6	0.7
Dorsal pectoral depth	27.3	26.6–30.7	28.6	1.7
Dorsal spine length	24.7	24.7–29.9	27.5	2.3
Dorsal pelvic depth	22.9	22.1–26.4	24.1	1.7
Dorsal-fin base length	31.2	29.1–31.2	30.0	1.0
Dorsal-adipose distance	11.2	10.5–13.7	11.6	1.2
Adipose-spine length	7.8	6.79–8.78	7.8	0.7
Dorsal adipose caudal distance	11.7	11.7–15.6	13.7	1.7
Caudal peduncle depth	12.5	12.5–14.2	13.3	0.6
Ventral adipose caudal distance	22.9	22.9–25.3	23.9	1.0
Adipose anal distance	21.3	18.5–21.3	19.8	1.0
Dorsal-anal distance	16.0	15.8–17.8	16.8	0.8
Pelvic-dorsal distance	29.5	22.0–29.5	22.5	2.7
Percents of head length (HL)				
Head-eye length	29.4	28.1–30.1	29.1	0.8
Orbital diameter	14.6	14.5–18.8	15.8	1.7
Snout length	63.2	63.2–70.5	66.8	3.1
Internares width	14.4	12.7–16.6	14.4	1.4
Minimal interorbital distance	28.8	28.8–35.7	32.2	2.5
Mouth length	53.8	52.0–60.6	55.7	3.5
Barbel length	14.0	7.6–14.0	10.6	2.6
Dentary tooth cup length	17.6	17.0–19.6	18.5	1.1
Premaxillary tooth cup length	17.8	17.2–19.2	18.2	0.7
Head depth	68.9	67.0–72.7	68.8	2.3

Burgos 2008 for comparison of both characters). Additionally, *P. zawadzki* differs from *P. nigrescens* by having rounded spots that do not cover more than one plate along the body (vs. whitish spots that become hazier along the body and can cover more than one plate, see *P. nigrescens* in fig. 3 in Chambrier and Montoya-Burgos (2008)). Moreover,

P. zawadzki is distinguished by having a greater head depth, 67.0–72.7% of HL (vs. 38.3–44.9% of HL in *P. barbatus*; 40.6–53.0% of HL in *P. corantijniensis*, data based on original description; and 52.5–56.6% of HL in *P. nigrescens*); greater cleithral width, 35.2–38.0% of SL (vs. 31.1–32.7% of SL in *P. nigrescens* and 29.7–33.4% of SL in *P. barbatus*); shorter distance between posteromedial margin of supraoccipital and origin of dorsal-fin, 6.7–9.2% of SL (vs. 10.4–11.6% of SL in *P. nigrescens*); greater anal-fin spine length, 11.9–13.8% of SL (vs. 7.3–10.4 of SL in *P. barbatus*); greater peduncle depth, 12.5–14.2% of SL (vs. 9.3–10.4 of SL in *P. barbatus*); and wider internares distance, 12.7–16.6% of HL (vs. 9.9–11.8% of HL in *P. barbatus*). *Pseudancistrus zawadzki* differs from *P. kwinti* and *P. guentheri*, two probable members of *P. barbatus* group by having whitish spots of the body (vs. body mottled or with bars, in *P. kwinti* and body plates dark at the base and pale along the edges, in *P. guentheri*).

Description. Morphometric data presented in Table 1. In lateral view, dorsal profile convex from snout tip to dorsal-fin origin; straight, gradually descending from dorsal-fin origin to posterior insertion of adipose fin; straight, steeply ascending to insertion of caudal fin; ventral profile flat from snout tip to anal-fin origin; shallowly concave from anal-fin insertion to lower caudal-fin spine; greatest body depth at dorsal-fin origin. In dorsal view, greatest body width across cleithral region; snout broadly elliptical; body progressively narrowed from opercular region to caudal fin. Cross-section of body between pectoral and pelvic fins rounded dorsally and flattened ventrally; cross-section of caudal peduncle ellipsoid.

Body almost entirely covered by plates; ventral portions of head and abdomen and dorsal-fin base naked. Five lateral rows of dermal plates, dorsal plates 21–24, lateral mid-dorsal plates 19–21, lateral median plates 22–24, lateral mid-ventral plates 21–24, lateral ventral plates 18–20. Three predorsal plates; eight plates below dorsal-fin base; four plates between dorsal fin and adipose fin; five rows of plates on caudal peduncle. Dorsal spinelet present.

Body plates and cleithrum have minute odontodes. Odontodes slightly hypertrophied on pectoral-fin spines, becoming gradually larger towards tips. Numerous yellowish hypertrophied odontodes along lateral margins of head including snout; odontodes small on tip of snout, increasing gradually in length from anterolateral margin of snout to cheeks; longest odontodes on posterior most portion of non-evertible cheek plates. Eyes small (orbital diameter 14.5–18.8% of HL), dorsolaterally positioned. Oral disk transversely ellipsoid. Lower lip not reaching transverse line between gill openings. Lower lip covered with numerous small papillae. Maxillary barbel developed. Mouth relatively large. Premaxillary teeth 40–61 per ramus; dentary teeth 28–69 per ramus. Teeth bifid, medial cusp large and rounded, lateral cusp minute and pointed. Wide jaws, dentary bones forming an oblique angle, premaxillary bones almost co-linear.

Dorsal fin II,7, origin approximately at midpoint between pectoral- and pelvic-fin origins, last dorsal-fin ray reaching adipose fin when depressed. Pectoral fin I,6, spine tip curved inward, covered with enlarged odontodes distally; depressed tip reaching one-third length of pelvic-fin spine. Pelvic fin I,5, spine tip curved inward, almost reaching anal-fin origin when depressed. Anal fin I,5, spine tip straight, reaching sev-



Figure 3. *Pseudancistrus zawadzki*, live specimen, LBP 15045, paratype, female, 128.7 mm SL, Tapajós river, Pará State, Brazil.

enth plate posterior to its origin. Caudal fin I,7–I,7, distal margin concave, inferior lobe longer than superior. Adipose fin with lightly curved spine, preceded by single median preadipose plate.

Color in life. Ground color dark greenish-brown on dorsum and sides of body, becoming dark brown posteriorly, and lighter brown ventrally. Anterior portion of head to posterior margin of orbits with many small, crowded, yellow spots; spots becoming abruptly larger on posterior portion of head, continuing on body, becoming slightly and gradually larger towards caudal peduncle. Dorsal plate series usually with two large spots per plate. Mid-dorsal plates usually with one large spot per plate. Lateral median plates with one large spot per plate. Mid-ventral plates and ventral plates with one large spot per plate. Dorsal-fin spine, rays and membranes with large round large spots. Adipose-fin with two large spots on spine and membrane. Pectoral, pelvic, anal and caudal fin with numerous and similarly sized yellow spots. Hypertrophied odontodes along head margin yellowish (Fig. 3).

Color in alcohol. Similar to pattern described for living individuals, but with ground color dark brown, and spots pale tan (Fig. 1).

Sexual dimorphism. Males possess a papilla posterior to urogenital opening, an attribute absent in females. Both sexes in *P. zawadzki* exhibit highly hypertrophied odontodes along snout margin, similar to others species of *Pseudancistrus* (Armbruster 2004b). In some loricariid species of genus *Pareiorhaphis* those hypertrophied odontodes may be sexually dimorphic (Pereira et al. 2007), an attribute not observed in the new species *P. zawadzki*.

Etymology. Specific name is in honor of Cláudio Henrique Zawadzki, professor at Universidade Estadual de Maringá (UEM), Maringá, Paraná State, Brazil, in recognition of his dedication and remarkable contributions to the study of the family Loricariidae.

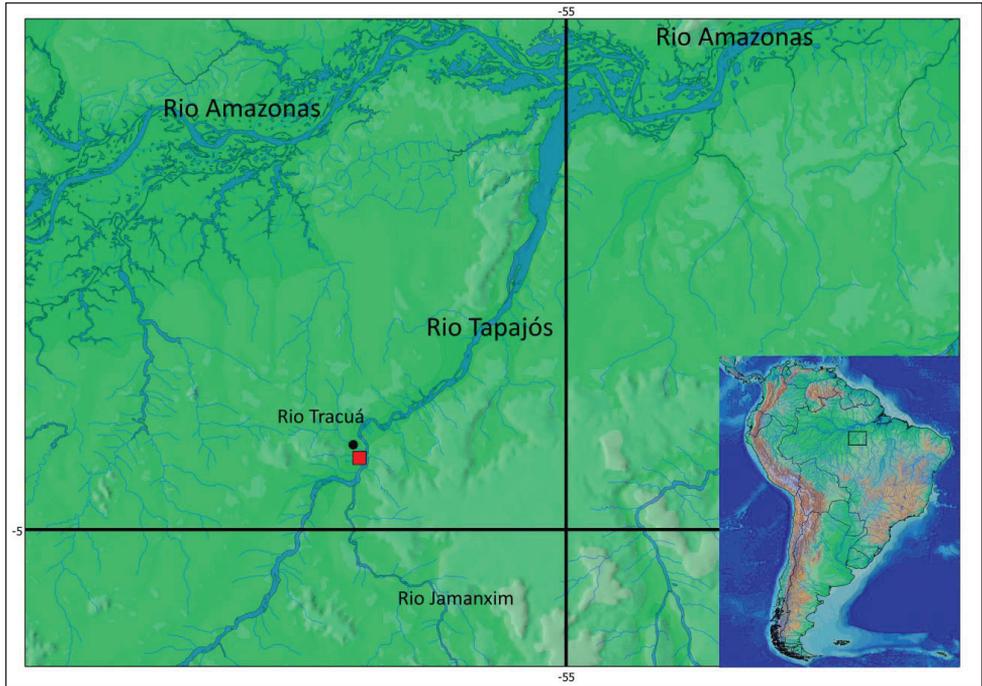


Figure 4. Map showing the type locality (red square) of *Pseudancistrus zawadzki* at rio Tapajós, $04^{\circ}33'09.7''\text{S}$, $56^{\circ}17'59.6''\text{W}$, and paratype locality (black circle) at rio Tracua, Tapajós river basin, $04^{\circ}28'11.2''\text{S}$, $56^{\circ}17'01.1''\text{W}$.



Figure 5. **a** Habitat at type locality of *Pseudancistrus zawadzki*: rio Tapajós, municipality of Itaituba, Pará State, Brazil **b** habitat at paratype locality: rio Tracua, Tapajós river basin, municipality of Itaituba, Pará State, Brazil.

Distribution. *Pseudancistrus zawadzki* is known from rio Tapajós ($04^{\circ}33'10''\text{S}$, $56^{\circ}18''\text{W}$) and rio Tracua ($04^{\circ}28'11''\text{S}$, $56^{\circ}17'01''\text{W}$), municipality of Itaituba, all from rio Tapajós basin, Pará State, Brazil. (see Fig. 4 for distribution map of type species localities).

Ecological notes. The rio Tapajós, and rio Tracuí where *P. zawadzki* occurs are clear water rivers, varying from medium to large size, with rocky outcrops forming small waterfalls and substrates of rocks and sand (Fig. 5).

Phylogenetic analysis

Partial sequences of the nuclear gene F-reticulon 4 (RTN4) were obtained in this study and from GenBank for 44 specimens representing 35 Loricariidae species and the new species *P. zawadzki* (Table 3). We included samples of the four lineages of *Pseudancistrus* proposed by Covain and Fisch-Muller (2012) to test whether *P. zawadzki* is part of the *P. barbatus* group. *Corydoras oiapoquensis* Nijssen, 1972 (Callichthyidae) was used to root the phylogeny. Additionally, samples of Delturinae (*Hemipsilichthys gobio* Lutken, 1874) and Loricariinae (*Harttia guianensis* Rapp Py-Daniel & Oliveira, 2001) were included in the analysis as additional outgroups. The combined sequence data resulted in a matrix with 2,318 base pairs (bp), out of which 1,079 were conserved and 896 were variable. The estimated index of substitution saturation (Iss) performed in DAMBE 5.2.31 (Xia and Xie 2001) showed that the data was not saturated (i.e. Iss.c value greater than Iss).

Evolutionary relationships among species of *Pseudancistrus* sensu lato and other members of Otothyriini are similar between our ML phylogenetic tree (-lnL = 11470.59) and the one proposed by Covain and Fisch-Muller (2012). In our analysis, the genus *Pseudancistrus* is paraphyletic with species assigned to three different lineages. The first lineage is monotypic, composed of *P. genisetiger*, sister to *H. gobio*, an outgroup taxon. Covain and Fisch-Muller (2012) suggested that *P. genisetiger* represents an undescribed genus within Delturinae. The second lineage of *Pseudancistrus* (*P. sidereus* + *P. pectegenitor*) is sister to a species of *Lithoxus* Eigenmann, 1910; Covain and Fisch-Muller (2012) suggested that the two species represent an undescribed genus or may be included in *Lithoxus*. The third lineage is composed of members of the *P. barbatus* group (*P. depressus*, *P. barbatus*, *P. corantijniensis*, *P. nigrescens*, the new species *P. zawadzki* and an undescribed species from the rio Xingu known as L17 among hobbyists). The *P. barbatus* group forms a polytomy with almost all species analyzed in the ingroup (Fig. 3), and was recognized by Covain and Fisch-Muller (2012) as true *Pseudancistrus* since this group includes the type species *P. barbatus*. Additionally, Covain and Fisch-Muller (2012) revalidated two genera for several species previously assigned to *Pseudancistrus*, - *Lithoxancistrus* (for *Pseudancistrus orinoco* (Isbrücker, Nijssen & Cala, 1988)) and *Guyanancistrus* (for *Pseudancistrus* sp., *P. brevispinis* (Heitmans, Nijssen & Isbrücker, 1983), *P. longispinis* (Heitmans, Nijssen & Isbrücker, 1983) and *P. niger* (Norman 1926)). Our analysis also supports the recognition and composition of those two genera.

Discussion

Taxonomy and phylogenetic comparison

The new species *P. zawadzkkii* possesses hypertrophied odontodes along the snout margin and lacks evertible cheek plates. Armbruster (2004b) identified that among Ancistrini, only *Pseudolithoxus*, *Lithoxancistrus*, and some members of *Guyanancistrus* and *Pseudancistrus* share the presence of hypertrophied odontodes along the snout in both sexes. Armbruster (2004b) also suggested that the species of *Pseudancistrus* that present this characteristic are derived; those species correspond to the *P. barbatus* group proposed by Chambrier and Montoya-Burgos (2008). Therefore, the new species described herein is a typical member of this group *sensu* Covain and Fisch-Muller (2012). Our phylogenetic analysis (Fig. 3) supports that hypothesis, and places the new species in a polytomy with *P. corantijniensis*, *Pseudancistrus* sp. L17 (undescribed species) and *P. nigrescens*, within the *P. barbatus* group. Our likelihood-based tests strongly rejected alternative topologies placing the new species in *Lithoxancistrus*, *Guyanancistrus* or with other species of *Pseudancistrus* apart from the *P. barbatus* group (see Table 2).

Pseudancistrus zawadzkkii, *P. corantijniensis*, and *P. nigrescens* share the presence of whitish colored snout odontodes and a dark colored body covered with white spots. The new species can be easily distinguished from *P. corantijniensis* and *P. nigrescens* by having large hypertrophied odontodes on the posteriormost portion of the non-evertible cheek plates, and marginal odontodes that increase gradually in length from tip of snout to cheeks. *Pseudancistrus barbatus* and *P. depressus* share reddish-brown snout odontodes, a probable synapomorphy, and are the sister group to *P. zawadzkkii*, *P. corantijniensis* and *P. nigrescens*. Covain and Fisch-Muller (2012) suggested that *P. guentheri* and *P. kwinti* may be added to the *P. barbatus* group. However, those two species have a different body coloration pattern (Chambrier and Montoya-Burgos 2008; see fig. 3); in *P. kwinti* the body is either mottled or with bars, while in *P. guentheri* the body plates are dark at the base and pale along the edges (Willink et al. 2010).

Biogeography and dispersal routes

Named species of the *P. barbatus* group are distributed in rivers draining to Guyana Shield into the Atlantic Ocean, and the new species described herein is from Tapajós river draining of Brazilian Shield into the Amazon. In our phylogeny, species from the eastern Guyana Shield (*P. barbatus* and *P. depressus*) form a clade sister to a group composed of species from the western Guyana Shield (*P. corantijniensis* and *P. nigrescens*) and Amazon basin (*P. zawadzkkii* and *Pseudancistrus* sp. L17) (Fig. 6). Therefore, based on this interpretation and our results of phylogenetic analysis, we suggested two hypotheses that could generate the distribution of *P. barbatus* group extant-species. The first hypothesis is that the ancestral stock of the *P. barbatus* group was widely distributed through all Guyana Shield rivers and Amazon Brazilian Shield rivers, and

Table 2. Likelihood-based tests for alternative topologies. SH and AU are probability values obtained from the Shimodaira-Hasegawa and the Approximately Unbiased tests (Shimodaira 2002). Asterisks denote significant values ($P < 0.05$ for SH and $P < 0.01$ for AU and ELW) that imply the topology is rejected.

Test	Topology	- Ln L	Δ - Ln L	ELW	SH	AU
	ML	11910.81				
1	<i>P. zawadzki</i> sister group to <i>P. pectegenitor</i> + <i>P. sidereus</i> ^a	11952.41	41.60	<0.001*	0.021*	<0.001*
2	<i>P. zawadzki</i> sister group to <i>Guyanancistrus</i> members ^a	11962.24	51.43	<0.001*	0.011*	<0.001*
3	<i>P. zawadzki</i> sister group to <i>Lithoxancistrus</i> members ^a	11966.25	55.44	<0.001*	<0.001*	<0.001*
4	<i>P. zawadzki</i> sister group to <i>P. genisetiger</i> ^a	12033.30	122.49	<0.001*	<0.001*	<0.001*

^a The alternative topology was defined as the ML tree forcing the desired relationship.

the species *P. zawadzki* and *Pseudancistrus* sp. L17 are in the limit of the distribution for the group in Tapajós and Xingu rivers, respectively. Gaston (1998) and Hubbell (2001) suggested that when allopatric divergence is the dominant mode of speciation, many daughter species are expected to arise from geographically widespread ancestral species. This is a reasonable interpretation given that named species of the group are widespread in rivers draining Guyana Shield into the Atlantic Ocean; the new species *P. zawadzki* are from Tapajós river drainage of Amazon Brazilian Shield; the possible new and undescribed species *Pseudancistrus* sp. L17 are from Xingu river which also belongs to drainages of Amazon Brazilian Shield and others possible new and undescribed species of *P. barbatus* group may be present in drainages of Guyana Shield into Amazon (*Pseudancistrus* sp. L220 from rio Paru; *Pseudancistrus* sp. L251 from rio Cuminá (rio Erepecuru); *Pseudancistrus* sp. L383 from rio Trombetas; *Pseudancistrus* sp. L440 from rio Jatapu (Seidel 2008)). However, phylogenetic and taxonomic studies are necessary to confirm that the latter undescribed species belong to *P. barbatus* group.

The second hypothesis suggests that the ancestral stock of *P. barbatus* group should have been distributed through Guyana Shield rivers and there existed several dispersal routes through Guyana and Amazon rivers, permitting that the ancestral lineages of *Pseudancistrus* sp. L17 and *P. zawadzki* reached the rivers of Amazon basin (see Fig. 7 for dispersal routes). Therefore, examples of connections and areas of movement among Guyana drainages and the north tributaries of Amazon basin was reported by several authors: (1) the Rupununi portal, an example of seasonal connection among Takutu and Rupununi rivers (Armbruster and Werneke 2005; Lujan and Armbruster 2011; De Souza et al. 2012); (2) the corridor among Sipalawini (Corantijn river basin) and the Paru do Oeste (Amazon basin), also connected only in the rainy season (Nijssen 1972; Lujan and Armbruster 2011); (3) the Cassiquiare Canal, a large and permanently navigable corridor between the upper Orinoco and the upper Rio Negro (Amazon) (Chernoff et al. 1991; Buckup 1993; Schaefer and Provenzano 1993; Lovejoy and Araújo 2000; Turner et al. 2004; Moyer et al. 2005; Willis et al. 2007; Winemiller et al. 2008; Winemiller and Willis 2011); (4) Proto-Berbice, a river system which had its headwaters in an ancient

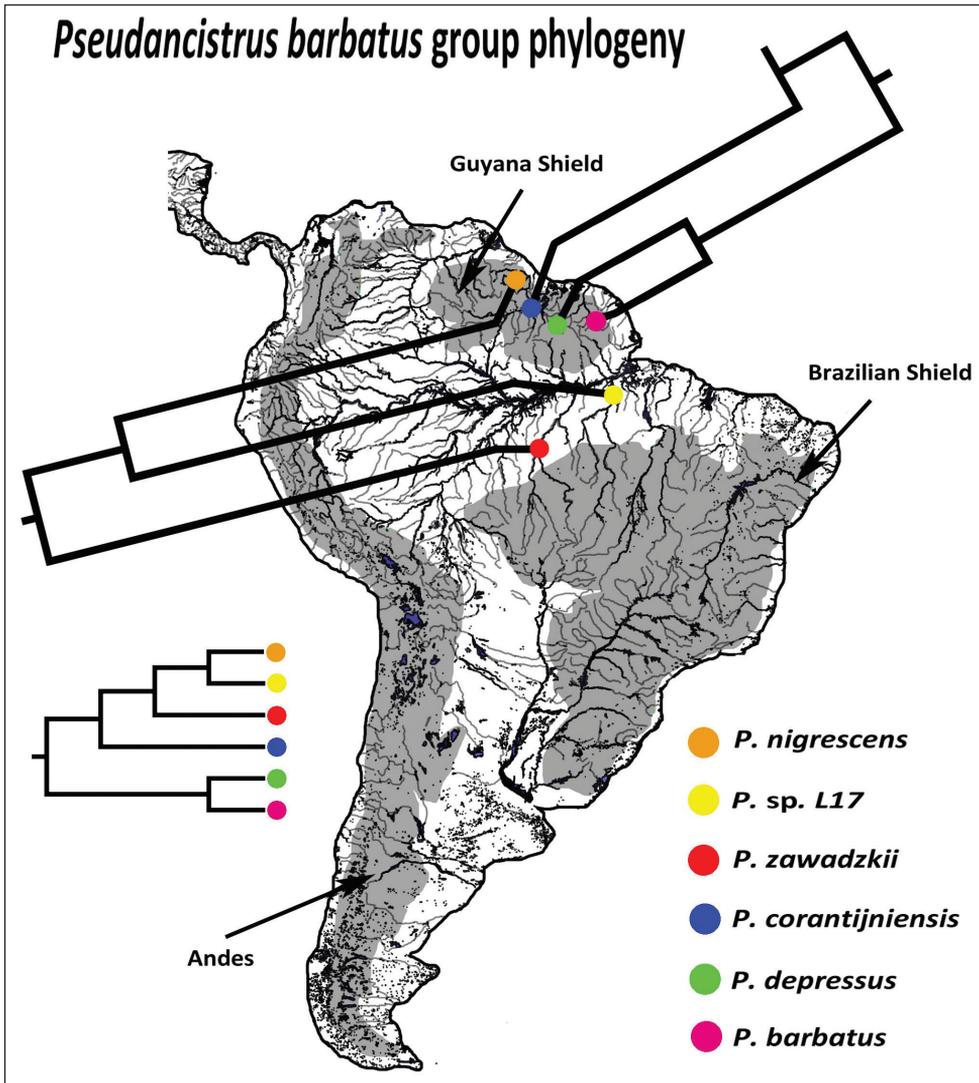


Figure 6. Distribution and phylogenetic relationships of species of the *Pseudancistrus barbatus* group based on F-reticulon 4 gene. Based in our first hypothesis of extend-species distribution of this group the ancestral was widespread through all Guyana Shield rivers and Amazon Brazilian Shield rivers, the species *Pseudancistrus zawadzki* and *Pseudancistrus* sp. L17 are in the limited distribution of this group in Tapajós and Xingu rivers, drainages of Brazilian Shield into Amazon.

mountain range draining northward to Guyana system (Rupununi and Essequibo rivers) and suffered a major sedimentation, erosion and/or corrosion of the highlands and at the end of the Pliocene had its head waters captured by the Amazon system; (5) the Atlantic coastal corridors resulted in a coastal marine corridor with reduced salinity due to the westerly Amazon River discharge, coastal junctions during times of marine regressions

Table 3. Taxa list, specimen and sequence data analyzed in the present study (n=44). Institutional acronyms follow Fricke and Eschmeyer (2013).

Species	Catalog Number	Field Number	GenBank N° F-RTN4	Ref.
<i>Corydonas oiapoquensis</i>	MHNG 2682.023	GF06-186	GU210997	Alexandrou et al. (2011)
<i>Hemipsilichthys gobio</i>	LBP 2368	15363	EU817547	Chiachio et al. (2008)
<i>Harttia guianensis</i>	MHNG 2643.016	GF00-351	FJ013232	Chiachio et al. (2008)
<i>Hypostomus</i> sp.	MHNG 2721.062	PE08-198	JN855790	Covain and Fisch-Muller (2012)
<i>Hypostomus boulengeri</i> (Eigenmann & Kennedy, 1903)	MHNG 2519.23	ASU7	EU817560	Chiachio et al. (2008)
<i>H. gymnorhynchus</i> (Norman, 1926)	MHNG 2621.098	SU01-160	JN855789	Covain and Fisch-Muller (2012)
<i>Ancistrus cirrhosis</i> (Valenciennes, 1836)	MHNG 2645.037	MUS 202	HM623638	Rodriguez et al. (2011)
<i>Dekeyseria picta</i> (Kner, 1854)	MHNG 2588.046	MUS 162	JN855755	Covain and Fisch-Muller (2012)
<i>D. scaphirhyncha</i> (Kner, 1854)	AUM 43874	V5528	JN855756	Covain and Fisch-Muller (2012)
<i>Hemiancistrus medians</i> (Kner, 1854)	MHNG 2664.078	GF00-084	JF747011	Fisch-Muller et al. (2012)
<i>Guyanancistrus brevispinis</i>	MHNG 2725.099	GF00-103	JN855772	Covain and Fisch-Muller (2012)
<i>G. brevispinis</i>	MHNG 2621.073	SU01-121	JN855773	Covain and Fisch-Muller (2012)
<i>G. longispinis</i>	MHNG 2725.100	GF99-204	JN855757	Covain and Fisch-Muller (2012)
<i>G. niger</i>	MHNG 2722.089	GF99-185	JN855759	Covain and Fisch-Muller (2012)
<i>Guyanancistrus</i> sp.	MHNG 2679.099	MUS 300	JN855774	Covain and Fisch-Muller (2012)
<i>Hopliancistrus tricornis</i> Isbrücker & Nijssen, 1989	MHNG 2588.051	MUS 146	JN855765	Covain and Fisch-Muller (2012)
<i>Lasiancistrus</i> aff. <i>caucanus</i>	MHNG 2586.043	MUS 118	JN855786	Covain and Fisch-Muller (2012)
<i>L. heteracanthus</i> (Günther, 1869)	MHNG 2613.037	CA 013	JN855787	Covain and Fisch-Muller (2012)
<i>L. planiceps</i> (Meek & Hildebrand, 1913)	STRI-01805	Stri 3526	JN855785	Covain and Fisch-Muller (2012)
<i>L. saetiger</i> Armbruster 2005	MHNG 2602.016	BR98-148	JN855754	Covain and Fisch-Muller (2012)
<i>L. schomburgkii</i> (Günther, 1869)	MHNG 2651.009	PE08-719	JN855782	Covain and Fisch-Muller (2012)
<i>L. schomburgkii</i>	MHNG 2651.068	GY04-308	JN855783	Covain and Fisch-Muller (2012)
<i>L. schomburgkii</i>	MHNG 2710.055	PE08-277	JN855784	Covain and Fisch-Muller (2012)
<i>L. tentaculatus</i> Armbruster, 2005	Mhng uncat.	MUS 573	JN855788	Covain and Fisch-Muller (2012)
<i>Lithoxus lithoides</i> Eigenmann, 1912	MHNG 2651.087	GY04-136	JN855777	Covain and Fisch-Muller (2012)
<i>L. pallidimaculatus</i> Boeseman, 1982	MHNG 2621.066	SU01-096	JN855778	Covain and Fisch-Muller (2012)
<i>L. planquettei</i> Boeseman, 1982	MHNG 2722.060	GF03-055	JN855779	Covain and Fisch-Muller (2012)

Species	Catalog Number	Field Number	GenBank N° F-RTN4	Ref.
<i>Lithoxancistrus orinoco</i>	AUM 43725	V5246	JN855766	Covain and Fisch-Muller (2012)
<i>L. orinoco</i>	AUM 42179	P4527	JN855767	Covain and Fisch-Muller (2012)
<i>Pseudancistrus barbatus</i>	MHNG 2653.059	GF00-074	JN855761	Covain and Fisch-Muller (2012)
<i>P. corantijiensis</i>	MHNG 2672.092	SU05-296	JN855781	Covain and Fisch-Muller (2012)
<i>P. depressus</i>	MHNG 2674.026	SU05-020	JN855780	Covain and Fisch-Muller (2012)
<i>P. genisetiger</i>	MHNG 2593.061	MUS 173	JN855764	Covain and Fisch-Muller (2012)
<i>P. nigrescens</i>	MHNG 2651.069	GY04-313	JN855770	Covain and Fisch-Muller (2012)
<i>P. nigrescens</i>	MHNG 2650.087	GY04-260	JN855771	Covain and Fisch-Muller (2012)
<i>P. pectegenitor</i>	AUM 42202	V5363	JN855769	Covain and Fisch-Muller (2012)
<i>P. pectegenitor</i>	ANSP 182801	V5433	JN855768	Covain and Fisch-Muller (2012)
<i>P. sidereus</i>	AUM 43443	P4871	JN855775	Covain and Fisch-Muller (2012)
<i>P. sidereus</i>	AUM 42180	P4537	JN855776	Covain and Fisch-Muller (2012)
<i>P. zawadzskii</i>	LBP 15045	61628	KJ028080	Present study
<i>Pseudancistrus</i> sp. L17	MHNG 2586.046	MuS 132	JN855763	Covain and Fisch-Muller (2012)
<i>Pseudolithoxus</i> cf. <i>kelsorum</i>	MHNG 2679.043	MUS 260	JN855762	Covain and Fisch-Muller (2012)
<i>P. dumus</i> (Armbruster & Provenzano, 2000)	MHNG 2708.080	MUS 288	JN855760	Covain and Fisch-Muller (2012)
<i>P. tigris</i> (Armbruster & Provenzano, 2000)	AUM 42215	V5292	JN855758	Covain and Fisch-Muller (2012)

and expanded coastal plains, and stream captures (Eigenmann 1912; Boeseman 1968; Cardoso and Montoya-Burgos 2009; Lujan and Armbruster 2011).

Additionally, the mainstream of Amazon River can act as a permeable barrier for endemic taxa on the respective Guiana and Brazilian shields. Several genera known to tolerate more lowland conditions (e.g. *Ancistrus* Kner, 1854, *Lasiancistrus*, and *Hypostomus* Lacepède, 1803) may be able to cross the Amazon basin, but such dispersal is unlikely among most species of Ancistrini (Lujan and Armbruster 2011). Also historically, epochs of cooler climate, as during glacial periods, could produce reduced precipitation, marine regressions, expansion of the coastal plain, and deepening of river channels. During such arid periods, rapids would have been more widespread, and deep-channel habitats that may currently work as barriers to fish dispersal would have been reduced (Schubert et al. 1986; Latrubesse and Franzinelli 2005; Lujan and Armbruster 2011). Drier climate will hardly change the Amazon river in a rapid, but can reduce its water flow allowing fish dispersal. Among Neotropical fishes *Psectro-*

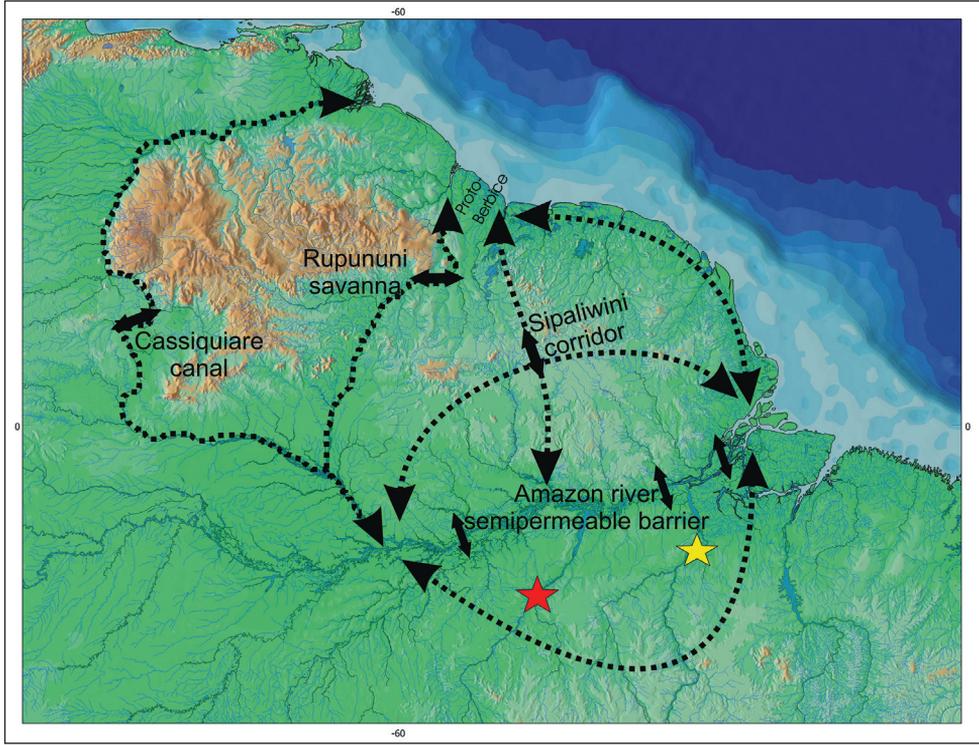


Figure 7. Hypothesized dispersal routs between basins of the Guiana Shield and Amazon Shield of ancestor of the *Pseudancistrus barbatus* group (based on Lujan and Armbruster 2011). Our second hypothesis of the *P. barbatus* group extent-species distribution is based on the assumption of a widespread ancestral through all Guyana Shield rivers and dispersal events enable the ancestor of *Pseudancistrus zawadzskii* (red star) and *Pseudancistrus* sp. L17 (yellow star) to colonize the Amazon Brazilian Shield rivers in Tapajós and Xingu rivers.

gaster essequibensis Günther, 1864 (Characiformes: Curimatidae; see Vari (1987)), *Parotocinclus aripuanensis* Garavello, 1988, and *P. britskii* Boeseman, 1974 (Loricariidae: Hypoptopomatinae) are species known to support dispersal via the northern Brazilian Shield.

Also, the dispersal routes around adjacent drainages of southern and northern Guyana Shield and northern parts of the Brazilian Shield could allow the dispersal of the ancestral form of *P. zawadzskii* and *Pseudancistrus* sp. L17, as well as others ancestral species of the *P. barbatus* group and even species of Ancistrini (Lujan and Armbruster 2011). The movement of fish species around adjacent drainages could be explained by two hydrographic reconfiguration process: headwater capture events (geomorphological phenomenon) and marine regressions (sea level oscillation). Changes in the earth's surface involving changes in the courses of rivers, as stream captures, portions of tributaries of a river in a watershed could be "captured" by adjacent basins resulting in isolated populations and at the same time letting species to move, or disperse, between

adjacent drainages (Almeida and Carneiro 1998; Bishop 1995; Wilkinson et al. 2006, 2010; Roxo et al. 2012). Montoya-Burgos (2003) hypothesized that dispersal (followed by allopatric population divergence) among Amazon and North-eastern coastal rivers probably occurred by temporary connections between adjacent rivers during periods of lower sea level about 6–5 Ma (see fig. 5 in Montoya-Burgos 2003). Cardoso and Montoya-Burgos (2009) suggested the same process to explain dispersal of *P. brevispinis* along coastal rivers of the Guyana. Therefore, temporary lowland connections and headwater capture events, together with the previously related hypothesis of colonization routes, likely explain the widespread distribution of the *P. barbatus* group extant species on Guyana and Brazilian Shields, as well as how the ancestral lineages of *P. zawadzki* and *Pseudancistrus* sp. L17 reached the drainages of the northern Brazilian Shield, in Tapajós and Xingu rivers.

Comparative material

Pseudancistrus barbatus (Valenciennes, 1840): ANSP 177366, 2, 76.5–103.7 mm SL, Burro Burro river, Water Dog Falls, Essequibo river basin, Guyana. ANSP 189119, 3, 75.1–151.5 mm SL, Lawa river, Sipalawini, Suriname. *Pseudancistrus brevispinis* (Heitmans, Nijssen & Isbrücker, 1983): ANSP 189128, 3, 56.8–125.7 mm SL, Marowini river, Sipalawini, Suriname. *Pseudancistrus nigrescens* Eigenmann, 1912: ANSP 177379, 5, 96.4–133.5 mm SL, Burro Burro river, Water Dog Falls, Essequibo river basin, Guyana. *Pseudancistrus orinoco* (Isbrücker, Nijssen & Cala, 1988): ANSP 160600, 6, 68.0–78.5 mm SL, Orinoco river, Venezuela. *Pseudancistrus pectegenitor* Lujan, Armbruster & Sabaj, 2007: ANSP 190755, 1, 206.2 mm SL, Ventuari river, Orinoco river basin, Venezuela. *Pseudancistrus sidereus* Armbruster, 2004b: ANSP 185321, 4, 148.6–154.1 mm SL, Casiquiari river, Venezuela. *Pseudancistrus* sp. L17: LBP 16551, 2, 75.3–101.0 mm SL; rio Xingu, Altamira, Pará State, Amazon river basin, Brazil. ANSP 193074, 3, 51.7–188.7 mm SL, Xingu river, Altamira, Pará State, Amazon river basin, Brazil. *Pseudancistrus* sp. ANSP 191153, 6, 49.2–75.7 mm SL, Ventuari river, Orinoco river basin, Venezuela.

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References

- Almeida FFM, Carneiro CDR (1998) Origem e evolução da Serra do Mar. *Revista Brasileira de Geociências* 28: 135–150.
- Armbruster JW, Provenzano F (2000) Four new species of the Suckermouth armored catfish genus *Lasiancistrus* (Loricariidae: Ancistrinae). *Ichthyological Exploration Freshwaters* 11: 241–254.
- Armbruster JW (2003) *Peckoltia sabaji*, a new species from the Guyana Shield (Siluriformes, Loricariidae). *Zootaxa* 344: 1–12.
- Armbruster JW (2004a) Phylogenetic relationships of the sucker-mouth armored catfishes (Loricariidae) with particular emphasis on the Hypostominae and the Ancistrinae. *Zoological Journal of the Linnean Society* 141: 1–80. doi: 10.1111/j.1096-3642.2004.00109.x
- Armbruster JW (2004b) *Pseudancistrus sidereus*, a new species from southern Venezuela (Siluriformes: Loricariidae) with a redescription of *Pseudancistrus*. *Zootaxa* 628: 1–15.
- Armbruster JW (2005) The loricariid catfish genus *Lasiancistrus* (Siluriformes) with descriptions of two new species. *Neotropical Ichthyology* 3(4): 549–569. doi: 10.1590/S1679-62252005000400013
- Armbruster JW (2008) The genus *Peckoltia* with the description of two new species and a reanalysis of the phylogeny of the genera of Hypostominae (Siluriformes: Loricariidae). *Zootaxa* 1822: 1–76.
- Bishop P (1995) Drainage rearrangement by river capture, beheading and diversion. *Progress in Physical Geography* 19: 449–473. doi: 10.1177/030913339501900402
- Boeseman M (1968) The genus *Hypostomus* Lacépède, 1803, and its Surinam representatives (Siluriformes, Loricariidae). *Zoologische Verhandelingen* 99: 1–89.
- Boeseman M (1974) On two Surinam species of Hypoptopomatinae, both new to science (Loricariidae, Siluriformes, Ostariophysi) *Proc. Kon. Nederl. Acad. Wetensch* 77(3): 275–271.
- Boeseman M (1982) The South American mailed catfish genus *Lithoxus* Eigenmann, 1910, with the description of three new species from Surinam and French Guyana and records of related species (Siluriformes, Loricariidae). *Proc. Kon. Nederl. Acad. Wetensch. (Ser. C, Biol. Med. Sci)*, 85(1): 41–58, pls. 1–5.
- Buckup PA (1993) Review of the characidiin fishes (Teleostei: Characiformes), with descriptions of four new genera and ten new species. *Ichthyological Exploration of Freshwaters* 4:97–154.
- Cardoso YP, Montoya-Burgos JI (2009) Unexpected diversity in the catfish *Pseudancistrus brevispinis* reveals dispersal routes in a Neotropical center of endemism: The Guyanas Region. *Molecular Ecology* 18: 947–964. doi: 10.1111/j.1365-294X.2008.04068.x

- Chambrier S, Montoya-Burgos JI (2008) *Pseudancistrus corantijniensis*, a new species from the Guyana Shield (Siluriformes: Loricariidae) with a molecular and morphological description of the *Pseudancistrus barbatus* group. *Zootaxa* 1918: 45–58.
- Chernoff B, Machado-Allison A, Saul WG (1991) Morphology, variation and biogeography of *Leporinus brunneus* (Pisces: Characiformes: Anostomidae). *Ichthyological Exploration of Freshwaters* 1: 295–306.
- Chiachio MC, Oliveira C, Montoya-Burgos JI (2008) Molecular systematic and historical biogeography of the armored Neotropical catfishes Hypoptopomatinae and Neoplecostomiinae (Siluriformes: Loricariidae). *Molecular Phylogenetics and Evolution* 49: 606–617. doi: 10.1016/j.ympev.2008.08.013
- Covain R, Fisch-Muller S (2012) Molecular evidence for the paraphyly of *Pseudancistrus sensu lato* (Siluriformes, Loricariidae), with revalidation of several genera. *Cybium* 36(1): 229–246.
- Cuvier G, Valenciennes A (1840) Histoire naturelle des poissons. Tome quinzième. Suite du livre dix-septième. *Siluroïdes* 15: 421–455.
- De Chambrier S, Montoya-Burgos JI (2008) *Pseudancistrus corantijniensis*, a new species from the Guyana Shield (Siluriformes: Loricariidae) with a molecular and morphological description of the *Pseudancistrus barbatus* group. *Zootaxa* 1918: 45–58.
- De Souza LS, Armbruster JW, Werneke DC (2012) The influence of the Rupununi portal on distribution of freshwater fish in the Rupununi district, Guyana. *Cybium* 36(1): 31–43.
- Eigenmann CH (1910) Catalogue of the fresh-water fishes of tropical and south temperate America. In: Reports of the Princeton University expeditions to Patagonia 1896–1899. *Zoology* 3: 375–511
- Eigenmann CH (1912) The freshwater fishes of British Guiana, including a study of the ecological grouping of species and the relation of the fauna of the plateau to that of the lowlands. *Memoirs of the Carnegie Museum* 5: 578 +51 pl.
- Eigenmann CH, Kennedy (1903) On a collection of fishes from Paraguay, with a synopsis of the American genera of cichlids. *Proceedings of Academy of Natural Sciences of Philadelphia* 55: 497–537.
- Eschmeyer WN, Fong JD (2013) Catalog of fishes. Online version, updated 07 January 2013. <http://research.calacademy.org/research/ichthyology/catalog/SpeciesByFamily.asp>
- Fowler HW (1941) A collection of fresh-water fishes obtained in eastern Brazil by Dr. Rodolpho von Ihering. *Proceedings of the Academy of Natural Sciences of Philadelphia* 93: 123–199.
- Fricke R, Eschmeyer WN (2013) Catalog of fishes. Online version, updated 07 January 2013. <http://research.calacademy.org/research/ichthyology/catalog/collections.asp>
- Garavello JC (1988) Three new species of *Parotocinclus* Eigenmann & Eigenmann, 1889 (Ostariophysi, Loricariidae) *Arquivos de Zoologia (São Paulo)* 28(4): 1–37. doi: 10.11606/issn.2176-7793.v28i4p1-37
- Gaston KJ (1998) Species-rangesize distributions: Products of speciations, extinctions and transformation. *Philosophical Transactions of the Royal Society of London, Series B-Biological Sciences* 353: 219–230. doi: 10.1098/rstb.1998.0204

- Günther A (1868) Diagnoses of some new freshwater fishes from Surinam and Brazil, in the collection of the British Museum. *Annals and Magazine of Natural History* 1(6): 475–481. doi: 10.1080/00222936808695733
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/92/NT. *Nucleic Acid Symposium Series* 41: 95–98.
- Heitmans WRB, Nijssen H, Isbrücker IJH (1983) The mailed catfish genus *Lasiancistrus* Regan, 1904, from French Guiana and Surinam, with descriptions of two new species (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde* 53(1): 33–48.
- Hubbell SP (2001) *The Unified Neutral Theory of Biodiversity and Biogeography*. University Press, Princeton, NJ.
- Isbrücker IJH, Nijssen H, Cala P (1988) *Lithoxancistrus orinoco*, nouveau genre et espèce de poisson-chat cuirassé du Rio Orinoco en Colombie (Pisces, Siluriformes, Loricariidae). *Revue française d'Aquariologie Herpétologie* 15: 13–16.
- Isbrücker IJH, Seidel I, Michels JP, Schraml E, Werner A (2001) Diagnose vierzehn neuer Gattungen der Familie Loricariidae Rafinesque, 1815 (Teleostei, Ostariophysi). *Datz-Sonderheft "Harnischwelse 2"*: 17–24.
- Jobb G, von Haeseler A, Strimmer K (2004) TREEFINDER: a powerful graphical analysis environment for molecular phylogenetics. *BMC Evolutionary Biology* 4: 4–18. doi: 10.1186/1471-2148-4-18
- Latrubesse EM, Franzinelli E (2005) The late Quaternary evolution of the Negro River, Amazon, Brazil: Implications for island and floodplain formation in large anabranching tropical systems. *Geomorphology* 70: 372–397. doi: 10.1016/j.geomorph.2005.02.014
- Lacepède BGE (1803) *Histoire naturelle des poissons*. Vol. 5. Plassan, Paris. 1xviii+803 p.+index, pls 1–21
- Lujan NK, Armbruster JW (2011) The Guiana Shield. In: Albert JS, Reis RE (Eds) *Historical Biogeography of Neotropical Freshwater Fishes*. University of California Press, Berkeley, 211–224. doi: 10.1525/california/9780520268685.003.0013
- Lujan NK, Armbruster JW, Sabaj MH (2007) Two new species of *Pseudancistrus* from southern Venezuela (Siluriformes: Loricariidae). *Ichthyological Exploration of Freshwaters* 18(2): 163–174.
- Lütken CF (1874) *Ichthyographische bidrag. I. Nogle nye eller mindre fuldstaendigt kjendte Pandsermaller, isaer fra det nordlige Sydamerica*. *Vidensk. Medd. Dansk Naturh. Foren. Kjob.* (for 1873), No. 13–14: 202–220, pl.4.
- Lovejoy NR, Araújo ML (2000) Molecular systematics, biogeography and population structure of Neotropical freshwater needle-fishes of the genus *Potamorhaphis*. *Molecular Ecology* 9: 259–268. doi: 10.1046/j.1365-294x.2000.00845.x
- Kner R (1854) Die Hypostomiden. Zweite Hauptgruppe der Familie der Panzerfische. (Loricata vel Goniodontes). *Denkschriften der Akademie der Wissenschaften in Wien*, 7: 251–286, pls. 1–5.
- Meek SE, Hildebrand (1913) New species of fishes from Panama. *Field Mus. Hist. Publ. Zool. Ser* 10(8): 77–91.

- Montoya-Burgos JI (2003) Historical biogeography of the catfish genus *Hypostomus* (Siluriformes: Loricariidae), with implications on the diversification of Neotropical ichthyofauna. *Molecular Ecology* 12: 1855–1867. doi: 10.1046/j.1365-294X.2003.01857.x
- Moyer GR, Winemiller KO, McPhee MV, Turner TF (2005) Historical demography, selection, and coalescence of mitochondrial and nuclear genes in *Prochilodus* species of northern South America. *Evolution* 59: 599–610.
- Nijssen H (1972) Records of the catfish genus *Corydoras* from Brazil and French Guiana with descriptions of eight new species (Pisces, Siluriformes, Callichthyidae). *Netherlands J. Zool.* 21(4): 412–433, pls. 1–3.
- Norman JR (1926) Descriptions of nine new freshwater fishes from French Guiana and Brazil. *Annals and Magazine of natural History.* (Ser. 9), 18 (no. 103): 91–97.
- Roxo FF, Zawadzki CH, Alexandrou MA, Costa Silva GJ, Chiachio MC, Foresti F, Oliveira C (2012) Evolutionary and biogeographic history of the subfamily Neoplecostominae (Siluriformes: Loricariidae). *Ecology and Evolution* 2: 2438–2449. doi: 10.1002/ece3.368
- Rapp Py-Daniel LH, Oliveira EC (2001) seven new species of *Harttia* from the Amazonian-Guyana region (Siluriformes: Loricariidae). *Ichthyological. Exploration of Fresh waters* 12(1): 79–96.
- Regan CT (1904) A monograph of the fishes of the family Loricariidae. *Transactions of the Zoological Society of London* 17 (3, no. 1): 191–350, pls. 9–21.
- Schaefer SA, Provenzano F (1993) The Guyana shield *Parotocinclus*: Systematics, biogeography, and description of a new Venezuelan species (Siluroidei: Loricariidae). *Ichthyological Exploration of Freshwaters* 4: 39–56.
- Schubert C, Briceño HO, Fritz P (1986) Paleoenvironmental aspects of the Caroni-Paragua River Basin (Southeastern Venezuela). *Interciencia* 11: 278–289.
- Seidel I (2008) Back to Nature Guide to L-Catfishes. Fohrman Aquaristik AB, Sweden, 208 pp.
- Shimodaira H (2002) An approximately unbiased test of phylogenetic tree selection. *Systematic Biology* 51: 492–508. doi: 10.1080/10635150290069913
- Shimodaira H, Hasegawa M (1999) Multiple comparisons of log-likelihoods with applications to phylogenetic inference. *Molecular Biology and Evolution* 16: 1114–1116. doi: 10.1093/oxfordjournals.molbev.a026201
- Stamatakis A, Hoover P, Rougemont J (2008) A rapid bootstrap algorithm for the RAxML web Servers. *Systematic Biology* 75: 758–771. doi: 10.1080/10635150802429642
- Strimmer K, Rambaut A (2002) Inferring confidence sets of possibly misspecified gene trees. *Proceedings of the Royal Society B* 269: 137–142. doi: 10.1098/rspb.2001.1862
- Turner TF, McPhee MV, Campbell P, Winemiller KO (2004) Phylogeography and intraspecific genetic variation of prochilodontid fishes endemic to rivers of northern South America. *Journal of Fish Biology* 64: 186–201. doi: 10.1111/j.1095-8649.2004.00299.x
- Wilkinson MJ, Marshall LG, Lundberg JG (2006) River behavior on megafans and potential influences on diversification and distribution of aquatic organisms. *Journal South American Earth Sciences* 21: 151–172. doi: 10.1016/j.jsames.2005.08.002
- Wilkinson MJ, Marshall LG, Lundberg JG, Kreslavsky MH (2010) Megafan environments in northern South America and their impact on Amazon Neogene aquatic ecosystems. In:

- Horn C, Wesselingh EP (Eds) Amazonia, Landscape and Species Evolution: A Look into the Past. Blackwell Publishing, 165–185.
- Willink PW, Mol JH, Chernoff B (2010) A new species of suckermouth armored catfish, *Pseudancistrus kwinti* (Siluriformes:Loricariidae) from the Copename River drainage, Central Suriname Nature Reserve, Suriname. *Zootaxa* 2332: 40–48.
- Willis SC, Nunes MS, Montana CG, Farias IP, Lovejoy NR (2007) Systematics, biogeography, and evolution of the Neotropical peacock basses *Cichla* (Perciformes: Cichlidae). *Molecular Phylogenetics and Evolution* 44: 291–307. doi: 10.1016/j.ympev.2006.12.014
- Winemiller KO, López-Fernández H, Taphorn DC, Nico LG, Barbarino-Duque A (2008) Fish assemblages of the Casiquiare River, a corridor and zoogeographical filter for dispersal between the Orinoco and Amazon basins. *Journal of Biogeography* 35: 1551–1563. doi: 10.1111/j.1365-2699.2008.01917.x
- Winemiller KO, Willis SC (2011) The Guiana Shield. In: Albert JS, Reis RE (Eds) *Historical Biogeography of Neotropical Freshwater Fishes*. University of California Press, Berkeley, 225–242.
- Valenciennes A (1836) Poissons [pl. 7]. In: A. d'Orbigny. *Voyage dans L'Amérique Méridionale (le Brésil, la République Argentine, la Patagonie, la République du Chili, la République de Bolivie)*.
- Xia X, Xie X (2001) Dambe: Data analysis in molecular biology and evolution. *Journal of Heredity* 92: 371–373.
- Xia X, Xia Z, Salemi M, Chen L, Wang Y (2003) An index of substitution saturation and its application. *Molecular Phylogenetic and Evolution* 26: 1–17. doi: 10.1016/S1055-7903(02)00326-3
- Xia X, Lemey P (2009) Assessing substitution saturation with DAMBE. In: Lemey P, Salemi M, Vandamme A-M (Eds) *The phylogenetic Handbook: A practical approach to DNA and Protein Phylogeny*. Second edition. Cambridge University Press, Cambridge, 615–630. doi: 10.1017/CBO9780511819049.022